

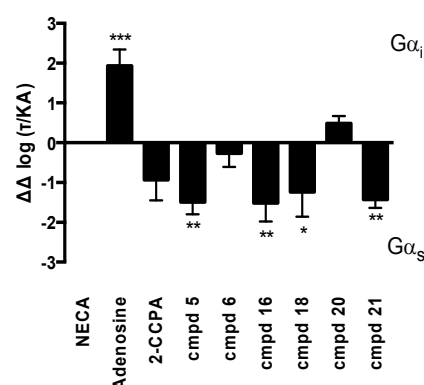
## Characterising signaling bias of novel Adenosine A<sub>1</sub> receptor agonists

The adenosine receptors (ARs) are family A G protein-coupled receptors (GPCRs) that exist as 4 subtypes; A<sub>1</sub>, A<sub>2A</sub>, A<sub>2B</sub> and A<sub>3</sub>, each with ~50-60% homology [1]. All four ARs modulate cAMP levels, with the A<sub>1</sub>R and A<sub>3</sub>R coupling to Gα<sub>i</sub> reducing cellular cAMP, while the A<sub>2</sub>Rs stimulate cAMP production via Gα<sub>s</sub> activation. All ARs are activated by the purinergic nucleotide, adenosine, and 2 synthetic adenosine derivatives; 5'-N-ethylcarboxamidoadenosine (NECA) and chloro-N<sup>6</sup>-cyclopentyladenosine (CCPA). We have developed novel AR ligands, some of which display A<sub>1</sub>R selectivity [2]. In an attempt to investigate which Gα<sub>i/o</sub> proteins couple to the A<sub>1</sub>R we used pertussis toxin (PTX), to inhibit the action of these, at the A<sub>1</sub>R. CHO cells, stably expressing the A<sub>1</sub>R (CHO-A<sub>1</sub>R), treated in this manner, were subsequently stimulated with our ligands, and cAMP levels measured. Interestingly we observed cAMP production upon PTX treatment (Table 1.), indicative of a Gα<sub>s</sub> component to the A<sub>1</sub>Rs repertoire. Whilst this has been observed before [3] it has not been seen to such significant levels, and has not been quantified. Thus here we present a quantification of the signaling bias between the canonical, Gα<sub>i</sub>, pathway, and non-canonical Gα<sub>s</sub> (Figure 1.). It is interesting to note that the ligand that displays the most extensive Gα<sub>i</sub> is the natural cognate ligand, adenosine. These agonists are currently the focus of on-going studies pertaining to; calcium mobilisation, ERK activation, and which inhibitory G proteins they activate.

**Table 1.** Potency and E<sub>max</sub> for CHO-A<sub>1</sub>R cells

Ligand	Gα <sub>i</sub> <sup>[2]</sup>		PTX (Gα <sub>s</sub> )	
	pEC <sub>50</sub>	E <sub>max</sub> <sup>a</sup>	pEC <sub>50</sub>	E <sub>max</sub>
NECA	-9.7 ± 0.2	59 ± 3.9	-6.7 ± 0.2***	44 ± 2.8*
Adenosine	-9.7 ± 0.3	59 ± 5.2	-4.7 ± 0.2***	58 ± 4.9
CCPA	-8.8 ± 0.2	69 ± 4.3	-7.0 ± 0.3***	38 ± 3.6***
cmpd 5	-6.9 ± 0.2	69 ± 4.3	-5.5 ± 0.3**	33 ± 5.3***
cmpd 6	-9.2 ± 0.2	56 ± 3.2	-6.7 ± 0.5***	26 ± 4.0***
cmpd 16	-7.7 ± 0.3	41 ± 4.8	-6.2 ± 0.3***	37 ± 4.1
cmpd 18	-8.7 ± 0.4	39 ± 4.7	-7.1 ± 0.3***	22 ± 2.8*
cmpd 20	-10.5 ± 0.3	48 ± 3.6	-7.1 ± 0.2***	40 ± 3.9
cmpd 21	-8.2 ± 0.2	47 ± 4.7	-6.7 ± 0.2**	47 ± 4.0

<sup>a</sup> – E<sub>max</sub> for Gα<sub>i</sub> coupling calculated as percentage inhibition when stimulated with 10μM forskolin. Statistical significance to Gα<sub>i</sub> calculated using a one-way ANOVA with Dunnett's post-test (\*, p < 0.05, \*\*, p < 0.01, \*\*\*, p < 0.001, \*\*\*\*, p < 0.0001)



**Figure 1. Relative bias for the A<sub>1</sub>R.** Signalling bias was calculated relative to NECA as ΔΔ log(τ/KA) [4]. Statistical difference to NECA was determined using a one-way ANOVA with a Bonferroni's post-test (\*, p < 0.05, \*\*, p < 0.01, \*\*\*, p < 0.001). Data are the mean of 5 individual data sets ± SEM.

[1] Jacobson and Gao (2006) *Nat Rev Drug Discov* **5**:247–264,

[2] Knight *et al* (2016) *J Med Chem* **59**:947-964,

[3] Baker and Hill (2007) *J Pharmacol Exp Ther* **320**:218–282,

[4] Baltos *et al* (2016) *Biochem Pharmacol* **99**:101–112.

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