

Rosiglitazone inhibits MMP-9 protein expression through NF-κB and AP-1 dependent mechanisms in human THP-1 cells

Omar Alshareif¹, Martina Hennessy¹, Matthew Hill², Joseph Keane³, J. Paul Spiers¹.

¹Department of Pharmacology and Therapeutics, Trinity College Dublin, Dublin, Ireland,

²Department of Psychiatry, Trinity College Dublin, Dublin, Ireland, ³Department of Respiratory Medicine, St James' Hospital, Dublin 8., Dublin, Ireland.

Altered expression of matrix metalloproteinases is associated with the development of atherosclerosis. Macrophages, a key component of the atherosclerotic plaque not only secrete MMP-9 but also express high levels of the peroxisome proliferators-activated receptor-γ (PPAR γ) nuclear receptor. Interestingly, glitazones (PPAR γ agonists) inhibit MMP-9 expression in macrophages and vascular smooth muscle cells. However, the underlying mechanism is less clear, with NF κ B, AP-1 and Stat pathways being implicated using luciferase reporter assay (Ricote *et al.* 1998). The aims of the present study were to investigate the direct effects of rosiglitazone (RGZ) and troglitazone (TGZ) on NF κ B, AP-1 and GSK-3B signalling with regard to the inhibitory effect of the glitazones on PMA-induced MMP-9 expression in THP-1 macrophages.

THP-1 cells were cultured in RPMI 1640 medium with 10% FBS and antibiotics. Cells were incubated in serum-free medium supplemented with medium (untreated control), or phorbol 12-myristate 13-acetate (PMA; 50ng/ml) for 48 hrs. RGZ or troglitazone (TGZ) were added to the medium prior to stimulation with PMA; cells were pre-exposed to GW9662 (PPAR γ antagonist) for 1hr where appropriate. MMP-9 expression was determined by ELISA and real time PCR. AP-1 transcription factors were analysed ELISA; NF κ B, I κ B α and GSK-3B were analysed by Western blot. Data were analysis by one-way ANOVA with post hoc analysis (Bonferroni) and expressed as mean \pm SE. A value of P<0.05 was taken to indicate statistical significance.

TGZ and RGZ inhibit PMA-induced MMP-9 mRNA and protein expression by 50 and 60% respectively. Furthermore, GW9662 reversed the inhibitory effect of RGZ (1 μ M) on MMP-9 protein expression (0.37 ± 0.02 vs 0.58 ± 0.036), but not mRNA expression; no effect was observed at higher concentrations of RGZ (10 μ M) or with TGZ. RGZ inhibited (P<0.05) PMA-induced NF κ B phosphorylation (2.0 ± 0.15 vs 1.4 ± 0.09), but did not affect I κ B α degradation after 48 hr exposure; no effect were observed at earlier time points. PMA alone or in combination with RGZ did not alter GSK-3B phosphorylation. RGZ inhibited PMA-induced expression of c-Jun, JunD, cFos, Fra1 and Fra2 members of the AP-1 transcription family by between 40-75%.

In conclusion, we demonstrate that RGZ and TGZ regulate MMP-9 protein and mRNA expression in THP-1 cells. In the case of RGZ this effect could be partially reversed by inhibition of PPAR γ . Furthermore, RGZ mediated inhibition of MMP-9 involved downregulation of NF κ B and multiple members of the AP-1 transcription family.

Ricote, M. *et al* (1998) *Natr.* **391**: 79-82.