

Heteromerisation with the bradykinin type 2 receptor enables β -arrestin recruitment to the angiotensin II type 2 receptor

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Bradykinin and angiotensin II are peptide hormones involved in the maintenance of blood pressure. Bradykinin reduces blood pressure via the bradykinin type 2 receptor (B₂R), while angiotensin II increases blood pressure primarily through the angiotensin II type 1 receptor (AT₁R). A second angiotensin II receptor (AT₂R) often, but not always, acts in opposition to the AT₁R, and its molecular and physiological functions remain poorly characterised [1,2]. Previous studies have demonstrated functional crosstalk between AT₂R and B₂R, with a suggested mechanism being heteromerisation [2].

Our aim was to provide evidence for the existence of the AT₂R-B₂R heteromer and investigate its pharmacology. HEK293FT cells were transfected with receptor cDNA fused to Rluc8 (variant of *Renilla* luciferase), untagged receptor, and β -arrestin2 fused to Venus (variant of green fluorescent protein). To investigate the existence of the AT₂R-B₂R heteromer, the G protein-coupled receptor-heteromer identification technology (GPCR-HIT) was employed on the bioluminescence resonance energy transfer (BRET) platform [3,4]. GPCR-HIT utilises ligand-dependent recruitment of interacting proteins such as β -arrestin2 to specifically detect receptor heteromers.

cDNA constructs were generated as described previously for other constructs [5]. AT₂R and V₂R were kindly provided by Walter Thomas (University of Queensland, Australia) and Thierry Durroux (Institute of Functional Genomics, Montpellier, France), respectively. B₂R was obtained from the Missouri S&T Resource Center. Transient transfections were carried out using GeneJuice (Merck, Kilsyth, Australia) and all BRET measurements were taken at 37°C.

Bradykinin treatment of cells co-expressing AT₂R-Rluc8, β -arrestin2-Venus and B₂R resulted in a ligand-dependent BRET signal (0.172 ± 0.011 , n=3) indicative of recruitment of β -arrestin2 to the heteromer. No signal was obtained when only AT₂R-Rluc8 and β -arrestin2-Venus were expressed, following treatment with either bradykinin (0.003 ± 0.007 , n=3) or angiotensin (0.0005 ± 0.006 , n=3). The GPCR-HIT signal was specific for the AT₂R-B₂R heteromer as the signal obtained when AT₂R-Rluc8 and β -arrestin2-Venus were co-expressed with the vasopressin type 2 receptor was significantly smaller (0.043 ± 0.020 , n=3, $p < 0.001$, two-way ANOVA compared to signal with AT₂R-Rluc8, β -arrestin2-Venus and B₂R). Subsequent antagonist assays showed that the bradykinin-induced BRET signal (0.153 ± 0.028 , n=3) could be significantly inhibited with a 30 minute pretreatment of the B₂R antagonist HOE140 (0.017 ± 0.025 , n=3, $p < 0.05$, one-way ANOVA) indicating that B₂R activation was required for recruitment of β -arrestin2 to the AT₂R-B₂R heteromer.

This study provides evidence for the existence of the AT₂R-B₂R heteromer. β -arrestin2 recruitment proximal to the AT₂R requires the specific co-expression and activation of the B₂R.

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