

Characterisation of the phospholipase A₂ isoforms supporting prostacyclin production synthesis by endothelial cells

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Endothelial cells line the luminal surface of blood vessels where they release hormones including prostacyclin (PGI₂) that dilates blood vessels, inhibits platelet aggregation and protects against atherogenesis. PGI₂ is produced by the sequential actions of three enzymes: phospholipase A₂ (PLA₂) cleaves membrane phospholipids to liberate arachidonic acid (AA), which is converted by cyclooxygenase (COX) into PGH₂, which is then converted to PGI₂ by PGI₂ synthase. Although numerous PLA₂ isoforms exist, it is thought to be the Ca²⁺-dependent cytosolic PLA₂ (cPLA₂) isoform that supports prostaglandin synthesis (1,2). However, it has been proposed that PGI₂ release from endothelial cells induced by thrombin or tryptase is dependent upon Ca²⁺-independent iPLA₂ (3). To test this proposal, we examined the contribution of cPLA₂ and iPLA₂ to PGI₂ synthesis using blood-outgrowth endothelial cells (BOECs) from healthy volunteers and from a cPLA₂-deficient patient (2), as well as intact mouse arteries.

BOECs were grown from blood of healthy donors (n=4) and a patient lacking cPLA₂ (2). These cells are an established model of endothelium from vessels (4). BOECs were incubated with the cPLA₂ inhibitor pyrrophenone (0.1-10μM) or vehicle (0.1% DMSO; 30min). They were then incubated with thrombin (1U/ml; PBS) or Ca²⁺ ionophore (A23187; 30μM; DMSO 0.1% final) to activate endogenous iPLA₂ and cPLA₂ enzymes, respectively, or with AA (50μM; ethanol 0.05% final), which stimulates PGI₂ formation without the requirement of PLA₂. In parallel, aortic rings prepared from mice (male, BALB/c, 10 weeks; n=6) were incubated with pyrrophenone, the iPLA₂ inhibitor bromoenol lactone (BEL; 0.1-10μM) or vehicle (0.1% DMSO), as above. Mouse aortic rings do not respond robustly to thrombin, so for these experiments, PGI₂ release was stimulated with acetylcholine (ACh; 1μM; saline). In this tissue ACh, as for thrombin, acts via a G_q-protein coupled receptor signalling pathway. In both assays, PGI₂ release was determined after 30min incubation by measurement of its breakdown product, 6-ketoPGF_{1α} using ELISA.

BOECs released PGI₂ in response to A23187, thrombin and AA (vehicle, 160±19pg/ml; thrombin, 1853±1089; A23187, 608±180pg/ml; AA, 2035±836pg/ml; all p<0.001 by Wilcoxon signed rank test). PGI₂ release induced by thrombin was inhibited in a concentration-dependent manner by the cPLA₂ inhibitor pyrrophenone (-logIC₅₀; 6.8±0.3, p=0.02 by Kruskal-Wallis test), as was release of PGI₂ induced by A23187 (-logIC₅₀; 7.1±0.7, p=0.05 by Kruskal-Wallis test). Release of PGI₂ from BOECs incubated with the COX substrate, AA, which bypasses the need for PLA₂ activity, was not affected by pyrrophenone. In agreement, BOECs from a cPLA₂-deficient individual did not produce PGI₂ in response to thrombin or A23187 but

responded normally to exogenous AA. In mouse aortic rings, ACh stimulated robust production of PGI₂ which was inhibited in a concentration-dependent manner by pyrrophenone (-logIC₅₀; 6.2±0.5; p=0.04 by Kruskal-Wallis test) but unaffected by the iPLA₂ inhibitor, BEL (ACh; 1374±252pg/ml, ACh+10μM BEL; 1377±201pg/ml, p=0.94 by Mann-Whitney test).

These data demonstrate that cPLA₂ and not iPLA₂ is the dominant isoform driving PGI₂ production by endothelial cells and intact arteries.

(1) Adler DH et al, JCI 118:2121, 2008. (2) Brooke MA et al, Gut *In press*, 2013 (3) Sharma J et al, Biochem 49:5273, 2010. (4) Starke RD et al, Blood, 121:2773, 2013.