

## Development of an ex vivo receptor occupancy assay for the Class C GPCR mGlu<sub>5</sub>

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**Introduction:** Receptor occupancy (RO) assays play an important role in drug development, as they confirm target engagement in the tissue of interest. We have recently developed HTL0014242, a negative allosteric modulator of mGlu<sub>5</sub>, and demonstrated RO in rat hippocampus using autoradiography<sup>1</sup>. However, such methodology requires specialised equipment and thus we have established an ex vivo RO assay using a tissue homogenate filtration-based format. Here, we report assay development and validation using [<sup>3</sup>H]M-MPEP and HTL0014242 in murine brain tissue.

**Method:** HTL0014242 was synthesized as established in<sup>1</sup>. Forebrain was dissected from six adult male CD-1 mice. Membranes were prepared from three mice as described previously<sup>2</sup> and protein linearity was performed to assess specific binding of [<sup>3</sup>H]M-MPEP. Using forebrain membranes (30 µg/well), saturation and kinetic [<sup>3</sup>H]M-MPEP binding experiments were performed. Competition binding assays were used to determine the pK<sub>i</sub> of HTL0014242. All were carried out as described in<sup>1</sup>. In RO experiments tissue needs to be rapidly processed to avoid dosed compound dissociation, therefore tissue was prepared by crude homogenisation (Polytron Homogeniser; 7,000 rpm; 20 s) immediately prior to use in protein linearity assays. Using chosen conditions (1.4 mg homogenate/well), the preparation was employed to perform both steady-state saturation and kinetic association assays at 4°C to select conditions resulting in full mGlu<sub>5</sub> RO.

**Results:** Increasing protein concentration correlated with increased specific binding of [<sup>3</sup>H]M-MPEP and 30 µg forebrain membranes/well resulted in high specific binding without radioligand depletion (Figure 1). Saturation and kinetic binding assays allowed K<sub>d</sub> and association/dissociation rates to be calculated, and competition binding assays established that HTL0014242 displays high affinity for the murine mGlu<sub>5</sub> (Table 1). By using high [<sup>3</sup>H]M-MPEP concentrations (20xK<sub>d</sub>) but short incubation times, it was demonstrated that all mGlu<sub>5</sub> receptors in tissue homogenates were occupied by 10 minutes, as calculated by comparing specific [<sup>3</sup>H]M-MPEP binding to the B<sub>max</sub> determined in steady-state saturation binding studies (Figure 2).

**Table 1.** mGlu<sub>5</sub> ligand binding in murine forebrain. K<sub>d</sub> of [<sup>3</sup>H]M-MPEP was determined by saturation binding (SB) or kinetic binding (KB) assays. Data shown are mean ± SD and as aim was to establish methodology, an n of 2 was performed to limit animal use.

[ <sup>3</sup> H]M-MPEP				HTL0014242
K <sub>d</sub> by SB (nM)	K <sub>on</sub> (M <sup>-1</sup> min <sup>-1</sup> )	K <sub>off</sub> (min <sup>-1</sup> )	K <sub>d</sub> by KB (nM)	pK <sub>i</sub>
3.91 ± 0.16	25,777,542 ± 2,477,593	0.0471 ± 0.0262	1.79 ± 0.84	8.88 ± 0.24

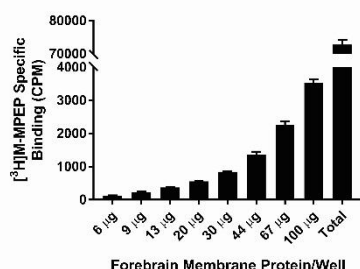


Figure 1. Binding of [<sup>3</sup>H]M-MPEP to membranes purified from murine forebrain

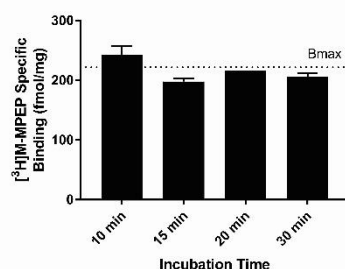


Figure 2. Association of 20xK<sub>d</sub> [<sup>3</sup>H]M-MPEP in crude homogenates of murine forebrain

**Conclusion:** Expression of mGlu<sub>5</sub> was confirmed in murine forebrain and binding of [<sup>3</sup>H]M-MPEP and HTL0014242 was characterised. In tissue from dosed animals, performing a 10-minute association of 20xK<sub>d</sub> [<sup>3</sup>H]M-MPEP at 4°C would allow the radioligand to occupy all unliganded mGlu<sub>5</sub> but should prevent

dissociation of HTL0014242. This data will serve as the groundwork for the final HTL0014242 dosing study to confirm mGlu<sub>5</sub> occupancy in the murine brain.

**References:**

1. Christopher JA *et al.* (2015). *J Med Chem.* **58**: 6653-64.
2. Bradley SJ *et al.* (2011) *Mol Pharmacol.* **79**: 874-85.