

## **A Chemical Characterisation Of Hydrogen Sulfide-Mediated Production Of Nitric Oxide From Nitrite And S-nitrosoalbumin**

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Until recently nitrite ( $\text{NO}_2^-$ ) has been considered just an inert metabolite of nitric oxide ( $\cdot\text{NO}$ ). In fact it is an important biological reservoir of  $\cdot\text{NO}$ .  $\text{NO}_2^-$  alters hypoxic signalling and vasodilation, and protects the vasculature after ischaemia-reperfusion injury.

S-nitrosothiols (RSNOs), another example of  $\cdot\text{NO}$ -derived species, have similarly gained great attention for their vasodilatory properties and vascular protective effects after ischaemic events. The mechanism suggested to explain both  $\text{NO}_2^-$  and RSNO-mediated effects is  $\cdot\text{NO}$  production.  $\text{NO}_2^-$  can be reduced to  $\cdot\text{NO}$  at low pH and low  $\text{O}_2$  tension by xanthine oxidase, deoxyhaemoglobin, mitochondrial complexes and ascorbate.

Hydrogen sulfide ( $\text{H}_2\text{S}$ ) is a recently discovered gasotransmitter which exerts many physiological effects similar to those of  $\cdot\text{NO}$  (e.g. vasodilation, promotion or inhibition of inflammation etc...)

The aim of this work was to test whether  $\text{H}_2\text{S}$  could promote  $\cdot\text{NO}$  production from  $\text{NO}_2^-$  and RSNOs. We also examined whether  $\text{H}_2\text{S}$  was able to interact with  $\text{NO}_2^-$  and RSNOs to affect cGMP production from human aortic smooth muscle cells (HASMC) and protect endothelial cells from oxidative stress-induced cytotoxicity.

$\cdot\text{NO}$  production from  $\text{NO}_2^-$  and S-nitrosoalbumin (SNOA) was chemically characterised by electron paramagnetic resonance (EPR) spin trapping and the spin trap used for the experiments was the  $\text{Fe}^{2+}$  complex of N-methyl-D-glucamine dithiocarbamate (MGD).  $\cdot\text{NO}$  was also assessed by gas phase ozone-based chemiluminescence. In the EPR experiments  $\text{H}_2\text{S}$  was generated using the sulfide salt NaSH. On addition of  $\text{H}_2\text{S}$  to  $\text{NO}_2^-$ , in the presence of the spin trap, a triplet EPR signal attributable to the  $\text{NO-Fe}^{2+}\text{-(MGD)}_2$  spin adduct was detected. Peak integration of the EPR spectra of spin-trapped  $\cdot\text{NO}$  showed that 5 min incubation (at room temperature) of 1 mM  $\text{NO}_2^-$  with 1 mM NaSH increased  $\cdot\text{NO}$  formation compared to  $\text{NO}_2^-$  alone, with increases of  $25.8 \pm 7.7\%$  (mean  $\pm$  SEM) and  $22.7 \pm 7.0\%$  at pH 4 and pH 7.4, respectively (n=4) (final  $\cdot\text{NO}$  concentrations in the presence of NaSH were  $579 \pm 67 \mu\text{M}$  and  $6.5 \pm 1.9 \mu\text{M}$  at pH 4 and 7.4 respectively). Chemiluminescence confirmed that  $\text{H}_2\text{S}$  rapidly reduced  $\text{NO}_2^-$  directly to  $\cdot\text{NO}$ , at pH 7.4 and that the reaction was thiol-dependent. Chemiluminescence also showed that, in the presence of NaSH, RSNO levels in SNOA significantly decreased (n=3,  $p < 0.01$ , data analysed by 2-way ANOVA) almost as soon as the reagents were mixed together.

This work sets the basis to assess whether  $\text{H}_2\text{S}$  mediated-reduction of  $\text{NO}_2^-$  and RSNOs can be considered a physiologically relevant alternative pathway for  $\cdot\text{NO}$  production.